

# Statistics for Biologists

## Lecture 2:

### Inferential statistics:

#### Pairwise comparisons, power and multiple testing

George Bell, Ph.D.  
Senior Bioinformatics Scientist  
Bioinformatics and Research Computing  
Whitehead Institute

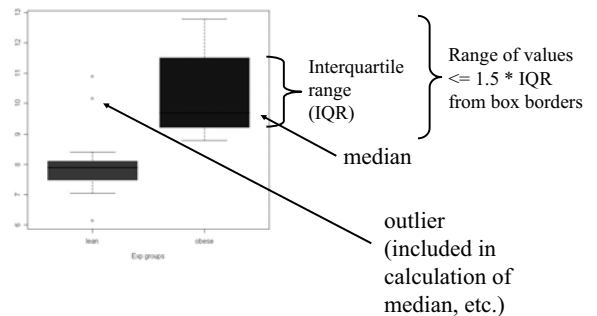
## Outline

- Lecture 1 review
- Intro to pairwise comparisons
- The t-test and other tests
- Statistical power
- Intro to multiple hypothesis testing
- Methods to correct for multiple hypothesis testing
- Using the R Commander
- Exercises

## From lecture 1

- Why use statistics?
- Descriptive statistics
  - central tendency + variability
- Visualization of quantitative data
  - What are you trying to show?
- Inferential statistics:  $H_0$ ,  $H_a$ ,  $\alpha$ ,  $\beta$
- False positives and false negatives
- Software for statistics
- Exercises

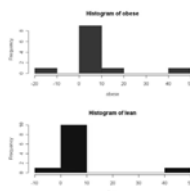
## Exercise 1 question: boxplot



## Exercise 1 questions: R

- Place multiple figures on the same plot:

```
par(mfrow=c(2,1))
hist(obese, col="red")
hist(lean, col="blue")
par() # reset
```



- Save a plot to a file: right click or
- ```
savePlot(filename="myPlot.pdf",
         device=2, type="pdf")
```

- Open another window for a new figure

```
windows()
```

## Intro to pairwise comparisons

- A common research question is,
  - “Is one measurement equal to another measurement?”
- More specifically, Q: “Is the mean of values in set X equal to the mean in set Y?”
  - sample answers: no; yes;  $X > Y$ ;  $X:Y = 1.5$
- To get an answer that includes some measure of confidence, we need to do a statistical test:
  - T-test (parametric: uses actual values)
  - Wilcoxon Rank Sum (Mann-Whitney) test (nonparametric: uses ranks; less powerful: greater  $\beta$ )

# Student's t-test

- History: Gossett ("Student") created this as a new technique to analyze Guinness beer using a small sample size.
- Assumptions:
  - measurements are independent
  - input data are normally distributed
  - Samples are from populations with equal variances (but test can be modified ("Welch's test") when this isn't true)
 Fortunately the test is robust, but...
- Two-tailed (testing for  $\neq$ ) or one-tailed (testing for  $>$  or  $<$ )
- T statistic = 
$$\frac{\text{mean}_1 - \text{mean}_2}{\text{SE}} \quad \text{df} = n_1 + n_2 - 2$$

# Performing the t-test

Data sets:  $a = \{ 2 \ 4 \ 6 \}$   $b = \{ 7 \ 8 \ 9 \}$

- Excel command: =TTEST(array1, array2, tails, type)  
example: =TTEST(A1:A3, B1:B3, 2, 3)
- R command: t.test(a, b, alternative="two.sided", var.equal=FALSE)

Output:

```
Welch Two Sample t-test
data: a and b
t = -3.0984, df = 2.941, p-value = 0.05479
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
-8.1553995  0.1553995
sample estimates:
mean of x mean of y
      4           8
```

Prob (H<sub>a</sub> is false)

# Comparing variances

- Use the F test
  - Excel: =FTEST(array1, array2)
  - R: var.test(a, b) # a=c(2, 4, 6); b=c(7, 8, 9)
- H<sub>0</sub>: True ratio of variances is 1

```
F test to compare two variances
data: a and b
F = 4, num df = 2, denom df = 2, p-value = 0.4
alternative hypothesis: true ratio of variances is not equal to 1
95 percent confidence interval:
0.1025641 156.0000000
sample estimates:
ratio of variances
      4 # To calculate variance: =VAR or 'var'
```

Prob (H<sub>a</sub> is false)

# The paired t-test

- Used when one measurement in each group comes from the same "experimental unit"
- example:
  - mouse assay after treatment by injection
  - one leg is treated; other leg is control
- Method (done by paired test):
  - get difference between each measurement pair
  - Are these differences  $\neq 0$ ?
  - (Do a one-sample t-test)
- Use when experimental design warrants it
- R command: t.test(a, b, paired=TRUE)

| Set A         | Set B |
|---------------|-------|
| 100           | 105   |
| 50            | 55    |
| 20            | 23    |
| 250           | 260   |
| 0.94 vs. 0.03 |       |

# T-test: summary

- Tests for a difference between two means
- Assumes data are normally distributed
- Test comes in multiple flavors:
  - 1 vs. 2 tails
  - equality of variances?
- Use the paired test when appropriate
- Excel just gives the take-home message
  - Or see Tools >> Data Analysis

# Moderated t-test

- Generic t statistic: 
$$t = \frac{\bar{X}_1 - \bar{X}_2}{s}$$
- Moderated t statistic: 
$$t = \frac{\bar{X}_1 - \bar{X}_2}{s + s_0}$$
  - Best known in microarray analysis
  - Corrects gene expression standard deviations toward a pooled estimate
  - Modify all standard deviations used in the t-test
  - $s_0 = 90^{\text{th}}$  percentile of all  $s$  (limma R package)
  - $s_0 =$  exchangeability constant (SAM)
  - gains power from sharing variation data across genes

## Power and sample size for the t-test

- Power reflects the probability of finding a true difference.
- Determinants of power:

$$n \geq \frac{2s_p^2}{\delta^2} (t_{\alpha, v} + t_{\beta(1, v)})^2$$

- Calculating power in R using  $\log_2$ -transformed data:
  - How many measurements are needed to be 80% sure that a 2-fold difference [ $\log_2(2)=1$ ] can be determined when using  $\alpha=0.05$  on data with a sd of 0.45 (standard deviation of  $\log_2$ -transformed measurements)?

```
power.t.test(n=NULL, delta=1, sd=0.45, sig.level=0.05, power=0.80,
type="two.sample", alternative="two.sided")
```

## Power and sample size for the t-test

R command: `power.t.test(n=NULL, delta=1, sd=0.45, sig.level=0.05, power=0.80, type="two.sample", alternative="two.sided")`

- Output:

**Two-sample t test power calculation**

```
n = 4.3824
delta = 1
sd = 0.45
sig.level = 0.05
power = 0.8
alternative = two.sided
```

**NOTE: n is number in \*each\* group**

## Wilcoxon Rank Sum test

- A non-parametric test:
  - An alternative to a t-test
  - Ranks of data (rather than actual values) are used.
- Also known as the Mann-Whitney test
- No assumptions about normally distributed data are required.
- Data sets
  - a = { 0.2 0.4 0.6 }      b = { 70 80 90 }
  - become      a = { 6 5 4 }      b = { 3 2 1 }
- R command:
 

```
wilcox.test(a, b, alternative="two.sided")      => p-value = 0.1
```
- But this test doesn't have much power:
  - same data in t-test yield a p-value of 0.005

## Intro to multiple hypothesis testing

- A p-value reflects the probability of a false positive call in a statistical test.
- When multiple hypotheses are tested on the same data, the rate of false positives greatly increases

Example for  $\alpha = 0.05$  :

| Number of genes tested (n) | Expected number of FPs = $\alpha * n$ | Probability of at least one FP = $100(1 - (1 - \alpha)^n)$ |
|----------------------------|---------------------------------------|------------------------------------------------------------|
| 1                          | 0.05                                  | 5%                                                         |
| 2                          | 0.1                                   | 9.75%                                                      |
| 100                        | 5                                     | 99.4%                                                      |

- So what to do?

## Choices for multiple hypothesis testing

- How many true positives do you expect?
- How costly is a false positive call?
- How costly is a false negative call?
- Major choices:
  - Family-wise error rate (ex: Bonferroni)
  - False discovery rate (ex: Benjamini and Hochberg)

## Corrections with the FWER

- Family-wise error rate = the probability of at least one false positive in the “family” of positive tests.
- “Bonferroni correction”
- A good choice if you predict that there are very few – if any – true positive tests
- Large correction means that
  - the false positive rate is very low
  - the false negative rate can be very high
- Practical implication: p-value must be really low to make it past a FWER correction.

## Performing a Bonferroni correction

- Corrected p-value = raw p-value \* n
  - n is the number of tests
  - if corrected p-value > 1, set to 1
- Example:
  - a microarray assays 10,000 genes
  - All are tested for differential expression with the t-test
  - If the raw p-value for one gene =  $10^{-5}$ 
    - The corrected p-value:
 
$$= 10^{-5} * 10,000 = 0.1$$
    - If  $\alpha = 0.05$ , then this gene cannot be described as differentially expressed.

## Corrections with the FDR

- False discovery rate = the frequency of false positives among the positive tests
- Introduced by Benjamini and Hochberg, 1995
- Tolerates a chosen proportion of false positives
- Much less conservative than Bonferroni:
  - false positive rate is higher
  - false negative rate is lower
- A good choice if you predict that there are many truly positive tests
- Practical implications:
  - more tests stay below  $\alpha$  compared to FWER
  - appropriate correction for many tests of differential expression in microarray experiments

## Performing a FDR correction

- Sort list of p-values in increasing order and starting at the bottom row
- Corrected p-value = the minimum between
  - raw p-value \* (n/rank)
  - corrected p-value below
  - n is the number of tests
  - rank is the position in the sorted list
- Example: a microarray assays 5 genes for differential expression

↑  
order of calculation

| Gene | Rank | Raw p-value | Formula                       | Corrected p-value |
|------|------|-------------|-------------------------------|-------------------|
| C    | 1    | 0.001       | $\min(0.001 * (5/1), 0.0125)$ | 0.005             |
| A    | 2    | 0.005       | $\min(0.005 * (5/2), 0.017)$  | 0.0125            |
| B    | 3    | 0.01        | $\min(0.01 * (5/3), 0.063)$   | 0.017             |
| E    | 4    | 0.05        | $\min(0.05 * (5/4), 0.1)$     | 0.063             |
| D    | 5    | 0.1         | $0.1 * (5/5)$                 | 0.1               |

## Performing multiple hypothesis corrections in R

```
# Read the data (tab-delimited text, with header fields; p-values in second field)
data.raw.pvals = read.delim("pvals_raw.txt", h=T)

# Correct the p-values using one of 7 methods ('?p.adjust' to see them)
# [,2] → use the data in column 2
FDR.p.vals = p.adjust(data.raw.pvals[,2], "fdr")

# Combine original file with corrected p-value output
data.adj = cbind(data.raw.pvals, FDR.p.vals)

# Print the output to a tab-delimited file
write.table(data.adj, file = "data_pvals_corrected.txt", sep="\t", quote=F)
```

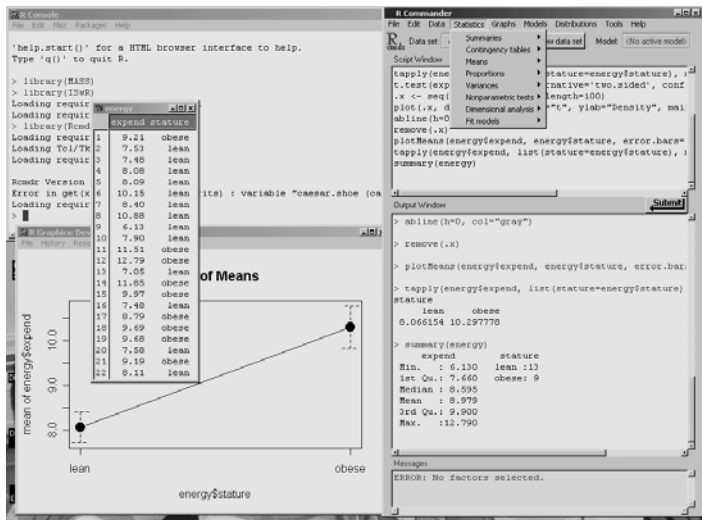
|   | id | Raw.p  | bonferroni | BY     | holm   | hochberg | BH     | fdr    |
|---|----|--------|------------|--------|--------|----------|--------|--------|
| 1 | A  | 0.0400 | 0.1600     | 0.1389 | 0.1200 | 0.1000   | 0.0667 | 0.0667 |
| 2 | B  | 0.0010 | 0.0040     | 0.0083 | 0.0040 | 0.0040   | 0.0040 | 0.0040 |
| 3 | C  | 0.2000 | 0.8000     | 0.4167 | 0.2000 | 0.2000   | 0.2000 | 0.2000 |
| 4 | D  | 0.0500 | 0.2000     | 0.1389 | 0.1200 | 0.1000   | 0.0667 | 0.0667 |

## Summary: multiple hypothesis corrections

- Beware that with raw  $\alpha = 0.05$ , 5% of tests will be positive just by chance
- Whenever performing >1 statistical test together, corrections should be done
- Select method based on desired FP and FN error rates
- Use R, Excel or BaRC web tool

## The R Commander

- Graphical interface designed to facilitate learning R
- Pull down menus and multiple-choice interface for common statistics and graphics
- Created by John Fox from McMaster University
- Installed with R on barra
- To start => begin R => library(Rcmdr)
- For Macs (but installation may be difficult), Windows, and Unix/Linux
- Other R graphical tools: affyImGUI, limmaGUI



## Summary

- Intro to pairwise comparisons
- The t-test and other pairwise tests
- Intro to multiple hypothesis testing
- Methods to correct for multiple hypothesis testing
- Using the R Commander
- Exercises

## References

- Zar JH. *Biostatistical Analysis*. Prentice Hall, 1998. [or any general biostatistics textbook]
- Dalgaard P. *Introductory Statistics with R*. Springer, 2002.
- Venables W.N. and Ripley B.D. *Modern Applied Statistics with S*. Springer, 2002.
- Tuft E. *The Visual Display of Quantitative Information*. Graphics Press, 1992.
- Lots of web sites
- R documentation

## Exercise 2 - To do

Using Excel and R:

- Transform data to get a more normal distribution
- Perform different t-test flavors for several different types of data
- Compute power for some t-tests
- Perform a Wilcoxon rank sum test
- Given a series p-values, perform multiple hypothesis testing
  - Bonferroni
  - FDR

## Helpful R commands

```

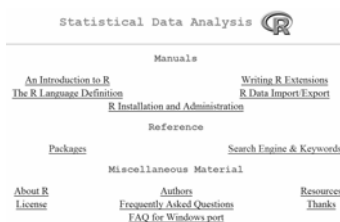
# HTML help page
help.start()

# Help for a command
?t.test

# Search help pages
help.search("t-test")

# Read a tab-delimited data table
energy = read.delim("energy.txt", header=T)
dat = read.delim(file("clipboard"), h=T)
rats = read.delim("http://.../rats.txt", h=T)

Google mailing lists, etc. => r-project t-test
  
```



## Exercise 2 functions

Excel

- TTEST
- RANK
- SORT
- LOG

R

- t.test
- wilcox.test
- p.adjust
- log
- power.t.test