

Bioinformatics for Biologists

Microarray Data Analysis. Lecture 1.

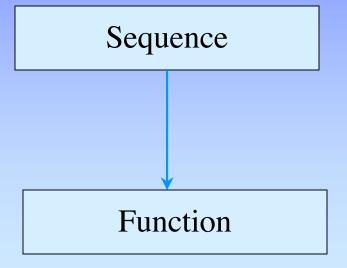
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Outline

- Introduction
- Working with microarray data
 - Normalization
 - Analysis
 - Distance metrics
 - Clustering methods

Research Trends

Genomics



- How are genes regulated?
- How do genes interact?
- What are the functional roles of different genes?
- How does expression level of a gene differ in different tissues?

Transcriptional Profiling

(Adapted from Quackenbush 2001)

- Study of patterns of gene expression across many experiments that survey a wide array of cellular responses, phenotypes and conditions
- Simple analysis what's up/down regulated?
- More interesting identify patterns of expression for insight into function, etc.

Microarray Data

Collect data on *n* DNA samples (e.g. rows, genes, promotors, exons, etc.) for *p* mRNA samples of tissues or experimental conditions (eg. columns, time course, pathogen exposure, mating type, etc)

Matrix (n x p) =

x_{11}	<i>x</i> ₁₂	• • • •	x_{1p}
<i>x</i> ₂₁	<i>x</i> ₂₂	• • • •	x_{2p}
•	•	•	•
x_{n1}	$x_{\rm n2}$	• • • •	$x_{\mathrm np}$

Multivariate Analysis

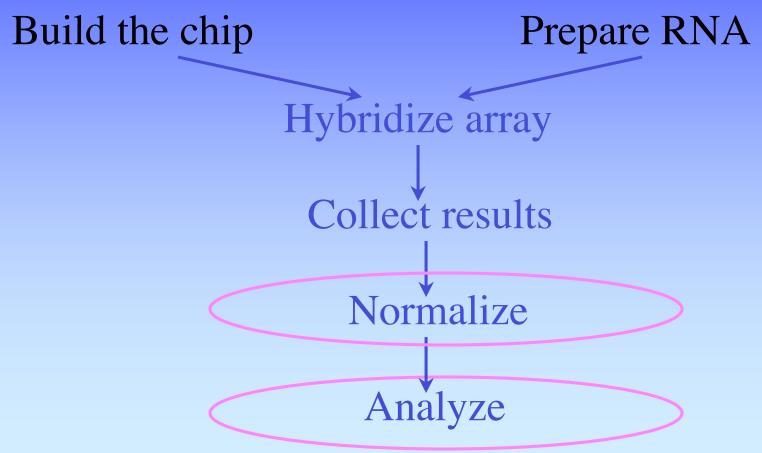
Concerned with datasets with more than one response variable for each observational or experimental unit (e.g. matrix X with *n* rows (genes) and *p* columns (tissue types))

- Hierarchical (phylogenetic trees) vs non-hierarchical (k-means)
- Divisive vs agglomerative
- Supervised vs unsupervised
 - Divide cases into groups vs discover structure of data

Multivariate Methods

- Cluster analysis discover groupings among cases of X
 - Hierarchical produces dendograms
 - K-means choose a prespecified number of clusters
 - Self Organizing Maps
- Principal component analysis (PCA)
 - Linear method, unsupervised, seeks linear combinations of the columns of X with maximal (or minimal) variance (graphical)

DNA Microarrays



Data Normalization

- Correct for systematic bias in data
 - Avoid it, recognize it, correct it, discard outliers
- First step for comparing data from one array to another

Sources of variation

wanted vs unwanted

Across experimental conditions

Chip, slide
Hybridization conditions
Imaging

Normalization Approaches

Compensate for experimental variability

- Housekeeping genes
- Spiked in controls
- Global median normalization
- Total intensity normalization
- LOWESS correction

Expression Ratios

- Let R = a query sample
- Let G = a reference sample
- Then the ratio, $T_i = R_i/G_i$
- Need to transform these to log₂
- Examples: T = 2/1 = 2; T=1/2 = .5
- Examples: $log_2(2) = 1$; $log_2(.5) = -1$

Total Intensity Normalization

(Adapted from Quackenbush 2002)

Assumptions: (1) start with equal amounts of RNA for the two samples; (2) arrayed elements represent random sample of genes in the organism

a.
$$N_{total} = \frac{\sum_{i=1}^{Narray} R_i}{\sum_{i=1}^{Narray} G_i}$$

c.
$$T_{i}' = \frac{R_{i}'}{G_{i}'} = \frac{1}{N_{total}} \frac{R_{i}}{G_{i}}$$

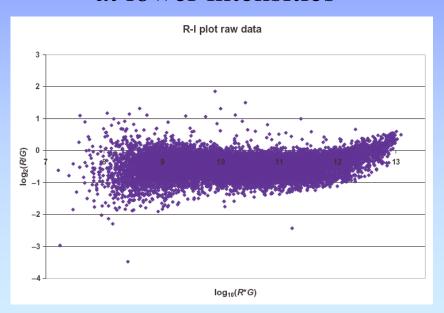
b. Rescale intensities:

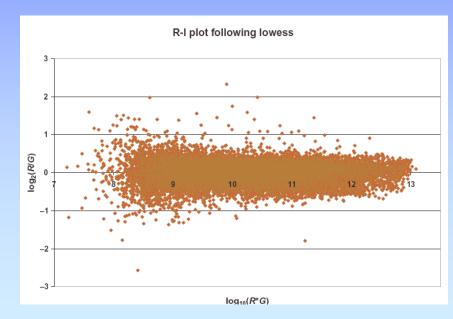
$$G'_i = N_{total}G_i$$
 and $R'_i = R_i$ ^{d.} $\log_2(T_i) \neq \log_2(T_i) - \log_2(N_{total})$

LOWESS - The R-I Plot

(Adapted from Quackenbush 2002)

- Data exhibit an intensity-dependent structure
- Uncertainty in intensity and ratio measurements is greater at lower intensities





LOWESS - The R-I Plot

(Adapted from Quackenbush 2002)

- Plot log₂(R/G) ratio as a function of log₁₀(R*G) product intensity
- Shows intensity specific artifacts in the measurements of ratios
- Correct using a local weighted linear regression

LOWESS Normalization

(From Quackenbush 2002)

If we set $x_i = \log_{10}(R_i * G_i)$ and $y_i = \log_2(R_i / G_i)$, lowess first estimates $y(x_k)$, the dependence of the $\log_2(\text{ratio})$ on the $\log_{10}(\text{intensity})$, and then uses this function, point by point, to correct the measured $\log_2(\text{ratio})$ values so that

$$\log_2(T_i) = \log_2(T_i) - y(x_i) = \log_2(T_i) - \log_2(2^{y(x_i)}),$$

or equivalently,

$$\log_2(T_i') = \log_2\left(T_i * \frac{1}{2^{y(xi)}}\right) = \log_2\left(\frac{R_i}{G_i} * \frac{1}{2^{y(xi)}}\right).$$

As with the other normalization methods, we can make this equation equivalent to a transformation on the intensities, where

$$G'_i = G_i * 2^{y(x_i)}$$
 and $R'_i = R_i$.

After normalization

(Adapted from Quackenbush 2001)

- Data reported as an "expression ratio" or as a logarithm of the expression ratio
- Expression ratio is the normalized value of the expression level for a particular gene in the query sample divided by its normalized value for the control

• Use log of expression ratio for easier comparisons

Citations

- Brazma A and Vilo J. Minireview: Gene expression data analysis. *FEBS Letters* 480:17-24, 2000.
- Quackenbush J. Computational Analysis of Microarray Data. *Nature Review* | *Genetics* 2:418-427, 2001.
- Quackenbush J. Microarray data normalization and transformation. *Nature Genetics Supp.* 32:496-501, 2002.
- Dudoit S and Gentleman R. Classification in microarray experiments. Statistics and Genomics Short Course -Lecture 5, January 2002 (http://www.bioconductor.org/workshop.html)

Lists of Tools

- Local WI Page
 - http://jura.wi.mit.edu/bio/microarrays/biopage5tools.html
 - WADE
- R Statistics Package Microarray Tools
 - http://www.stat.uni-muenchen.de/~strimmer/rexpress.html
- Bioconductor Project
 - http://www.bioconductor.org/
- EBI
 - http://ep.ebi.ac.uk/Links.html
 - http://ep.ebi.ac.uk/EP/

Exercise 1

Excel Conventions

• A2

• A2:A100

• =B5

• =\$B\$5

• =data!B4

• =[otherFile.xls]data!B4 reference other file

cell reference

series of cells

formula

absolute link

reference other sheet

Exercise 1 Functions

- MEDIAN
- SUM
- AVERAGE
- IF
- TTEST
- VLOOKUP

Exercise 1 To Do

Affy - fetal & human adult liver & brain tissue

- Normalize data 8 chips (replicates)
 - Global median normalization
 - (expression signal/chip median value)*100
- Filter low intensity signals
 - Based on A/P
 - Eliminate signal similar to background
- Calculate ratios
 - Reduce data (replicates)
 - Use AVERAGE function
 - Ratio of fetal tissue/adult tissue
 - Log₂